

# Biochemical Stability and Interactions of Dental Resin Composites and Adhesives with Host and Bacteria in the Oral Cavity: A Review

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# Abstract

The use of resin composites and adhesives in dental restorations is ubiquitous. However, the longevity of resin composites is less than that of comparable restorative materials, mainly because of higher fracture rates and greater prevalence of secondary caries. Dental resin composites and adhesives contain ester links, which are vulnerable to biochemical hydrolysis by esterase activity from human saliva and bacteria. In this article, we review biodegradation processes that occur in the oral cavity and their contribution to the premature failure of resin composites. Biodegradation causes deterioration of resin composite bulk and the composite-tooth interface and releases by-products, such as methacrylic acid, triethylene glycol and bishydroxy-propoxy-phenyl-propane. These by-products have been shown to affect cariogenic bacterial growth and virulence. A compromised restoration-tooth interface allows saliva and oral bacteria to infiltrate the spaces between the tooth and the composite, exacerbating the effects of biodegradation, undermining the restoration and leading to recurrent caries, hypersensitivity and pulpal inflammation. It is important to consider the biochemical stability of these materials to advance their chemistry beyond the current formulations and conceive more biochemically stable and better-performing dental resin composites and adhesives.

ver the last several decades, concerns about the possibility of adverse health effects from exposure to mercury in dental amalgams and the desire for improved esthetic dental restorations have led to a steady and rapid increase in the use of resin composite restorations.<sup>1-3</sup> Since the late 1990s and early 2000s, there has been a strong trend in terms of decreased use of amalgams, countered by an increase in the use of resin composites.<sup>4</sup> However, dental resin composites have an average replacement time of 5.7 years, mainly because of secondary caries and fracture of the restoration (**Fig. 1**).<sup>5</sup> Therefore, despite the esthetic appeal and reduced preparation size of the restoration associated with resin composites,<sup>6</sup> concerns over higher fracture rates, reduced longevity, prevalence of secondary caries and bacterial proliferation have emerged.<sup>2,7,8</sup>

Most research on resins has focused on physical processes that lead to degradation<sup>9</sup>; however, recently, attention to biochemical processes leading to degradation has increased.<sup>10-12</sup> Resin composites contain ester

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Figure 1: Radiographic image of secondary caries at the resin compositetooth interface (arrows).

links that are vulnerable to hydrolysis by esterase activity in the oral cavity.<sup>10,13-17</sup> Biodegradation results in deterioration of the bulk structure of resin composites as well as the composite-tooth interface and the release of degradation products, such as methacrylic acid, triethylene glycol (TEG) and bishydroxy-propoxy-phenyl-propane (BisHPPP) (**Fig. 2**).<sup>10,12,17</sup> These products have been shown to affect cariogenic bacterial growth and virulence.<sup>7,18-21</sup> The compromised composite-tooth interface allows saliva and bacteria to infiltrate the spaces between the tooth and composite, exacerbating the effects of biodegradation, undermining the restoration and likely contributing to recurrent caries, hypersensitivity and pulpal inflammation.<sup>22-27</sup>

This review begins with an introduction to resin composites and adhesives, followed by an analysis of the biochemical stability of these materials when challenged with human salivary esterases. The effects of bacteria on the biochemical stability of resin composite restorations and the reciprocal effects of degradation by-products on the bacteria are then discussed.

# **Resin Composites**

The 4 constituents of dental resin composites are a polymeric matrix (usually methacrylate based); filler particles (usually glass, quartz or ceramic oxide, such as alumina or silica); coupling agents (used to improve bonding of the filler–polymer matrix); and a photoinitiator–inhibitor system.<sup>9,11,12,28</sup>

## The Matrix

Dental resin monomers, which are the building blocks of

the polymeric matrix, have been based primarily on the coupling of chemical moieties via ester links (**Fig. 2**). Some of the most popular resin monomers are bisphenol-glycidyl-dimethacrylate (BisGMA), triethylene glycol dimethacrylate (TEGDMA), urethane dimethacrylate (UDMA) and bisphenol A polyethylene glycol diether dimethacrylate (BisEMA) (**Fig. 2**).<sup>9,12,28</sup>

BisGMA is probably the most widely used monomer in both resin composites and adhesives. Its popularity is attributed to its relative non-volatility, low degree of polymerization shrinkage, rapid hardening under oral conditions and compatibility with current inorganic filler systems.<sup>9,12,28</sup> A disadvantage of BisGMA is its high viscosity, which results from hydrogen bonds between hydroxyl groups in the alkyl chains and a rigid aromatic ring structure (**Fig. 2**). To facilitate handling and manipulation, various diluent monomers are used in conjunction with BisGMA, most commonly TEGDMA (**Fig. 2**), but other monomers, such as UDMA and BisEMA, are also used.<sup>9,12,28</sup> The ratios and compositions of the monomers constituting resin composites vary depending on the application, location (anterior vs. posterior) of the restoration and the manufacturer's preference.<sup>9,12,28</sup>

### Filler Systems

Reinforcing fillers are major constituents of resin composites by weight and volume (about 50–80%). Fillers improve the physical properties of the composite, such as strength and modulus of elasticity, as well as reducing polymerization shrinkage, the coefficient of thermal expansion and water sorption.<sup>9,11,28,29</sup>

## **Coupling Agents**

Coupling agents have been developed to increase the strength of the chemical link between the hydrophobic organic matrix and hydrophilic inorganic fillers. They have hydrophilic hydroxyl groups at one end and hydrophobic methacrylate groups at the other (**Fig. 3**). The most common coupling agents are organic silicon compounds called silanes.<sup>9,11,28</sup>

#### **Polymerization Systems**

Composites are converted from a viscous resin to a rigid solid through free radical polymerization of the methacrylate monomers. During polymerization, each molecule grows by the addition of a monomer to a terminal free radical reaction site. However, not all of the monomer's double bonds will have reacted once polymerization has stopped. In fact, studies suggest that the degree of vinyl group conversion of the methacrylate materials ranges from 40% to 85%.<sup>11-13,30</sup> The incomplete conversion is, in part, a result of a reduction in the diffusion rate of the propagating free radicals, the unreacted dimethacrylate molecules and the pendant methacrylate species as polymerization progresses.<sup>11,12,17</sup> Thus, following the polymerization process, a significant amount of partly reacted and unreacted





Bis-hydroxy-propoxy-phenyl-propane (BisHPPP) Methacrylic acid (MA)

Β.



Triethylene glycol dimethacrylate (TEGDMA)



Triethylene glycol (TEG)

**Figure 2:** Structure and hydrolysis of the common dimethacrylate monomers, bisphenol-glycidyl-dimethacrylate (BisGMA) (a) and triethylene glycol dimethacrylate (TEGDMA) (b), by salivary and bacterial esterases, resulting in the production of the by-products, bishydroxy-propoxy-phenyl-propane (BisHPPP), triethylene glycol methacrylate (TEGMA), triethylene glycol (TEG) and methacrylic acid (MA).

monomers remains in the polymeric matrix. Unreacted hydrophilic monomers may diffuse into saliva and interact with salivary proteins, bacteria and host tissue.<sup>7,10-14,16-21,26,31-34</sup>

#### **Dental Resin Adhesives**

Resin composite restorations require the application of

adhesives to bond them efficiently to a tooth (specifically dentin). Dental resin adhesives are low-viscosity methacrylate-based liquids, which spread on the dentin surface and solidify to bond to the primed dentin on one side and to the resin composite substrate on the other.<sup>25,35</sup> Dental adhesives and primers contain resin monomers that are similar to those found in the composite resin matrix by the inclusion of methacrylate functional groups, yet they contain hydrophilic moieties, such as hydroxyethyl methacrylate,<sup>25</sup> to allow proper interactions with the tooth material. These adhesive monomers provide the covalent link between the adhesive and the composite, thus ensuring structural continuity and physical co-mechanical properties, such as strength. Currently, 2 types of adhesive systems are in use: total-etch (also known as etch-and-rinse) and self-etch systems.<sup>25</sup>

#### **Total-Etch Adhesives**

The first step in total-etch systems is the demineralization of the dentin surface to a depth of up to 20 µm by the application of an acid (usually 30–40% phosphoric acid), followed by a rinsing step and then the application of the primer and adhesive agents, which will infiltrate the exposed collagen and polymerize to form the resin–dentin interface or "hybrid layer."<sup>25,36,37</sup> The application of a primer followed by the adhesive agent successively in separate steps forms the basis for the 3-step etch-and-rinse technique. Simplified 2-step etch-and-rinse adhesives combine the primer and adhesive agent in 1 step.<sup>37</sup>

#### Selt-Etch Adhesives

A trend toward simpler techniques that are less practitioner sensitive and more time efficient has resulted in the manufacture of self-etch adhesives.<sup>25</sup> These adhesives use non-rinse acidic monomers that simultaneously condition and prime dentin to form the hybrid layer. Self-etch adhesives may be 2-step or 1-step, depending on whether the hydrophobic adhesive resin is applied in a separate step or combined with the hydrophilic self-etch primer, respectively.<sup>38</sup>

## Biodegradation of Dental Resin Composites and Adhesives

The degradation of dental resin composites and adhesives in the oral cavity is a result of many complex interactions,





Figure 3: Structural formula of  $\gamma$ -methacryloxypropyltrimethoxy silane.

physical and biochemical. Most research has focused on the physical processes (food, chewing, etc.) that lead to degradation. These are classified as either material loss and uptake (sorption, extraction, dissolution and mineralization) or physical changes (softening, stress cracking, fatigue fracture, etc.).<sup>9</sup> In contrast, biochemical processes have seldom been discussed until recently.

Biochemical processes that lead to degradation are thermolysis (decomposition caused by heat), oxidation (loss of electrons), solvolysis (decomposition caused by a solvent), photolysis (decomposition caused by light) and radiolysis (decomposition by ionizing radiation). Solvolysis, and more specifically hydrolysis when the solvent is water, is the most investigated and most relevant biochemical degradation process as it acts on the unprotected ester linkages in methacrylate-based resin monomers, polymers and coupling agents. By definition, hydrolysis is a chemical reaction during which water cleaves a molecule into two parts. One fragment of the parent molecule gains a hydrogen ion and the other a hydroxyl group from the water molecule (**Fig. 2**).<sup>9,10,12,17,31</sup>

The potential for enzymes to interact with resin composites and adhesives is significant. In the oral cavity, the enzymes most associated with the hydrolysis of resin composites and adhesives are esterases. Salivary esterases originate in the human gingival epithelium and salivary glands, through inflammatory responses and from microorganisms.<sup>9,10,16,17,32,33</sup>

Studies have shown the ability of human saliva to hydrolyze dental resin monomers, as well as the polymerized matrix.<sup>10,16,23,26,33,34,39</sup> For example, Jaffer and colleagues<sup>17</sup> showed that human saliva degrades the commercial composite resins Filtek Z250 (3M ESPE, St. Paul, MN, USA) and Spectrum TPH (L.D. Caulk, Milford, Del., USA), both of which contain BisGMA, TEGDMA and urethane-modified BisGMA<sup>12</sup>. In this study, standardized commercial photopolymerized composites were incubated with human saliva for up to 16 days (pH 7.0 and 37°C). Analysis of the incubation solutions revealed that human saliva catalyzed the biodegradation of both composites in a materials-dependent

manner. Biodegradation products included methacrylic acid, TEGMA, BisHPPP and ethoxylated bisphenol A (E-BPA). The latter is a by-product of BisEMA biodegradation.<sup>12</sup>

The nature of degradation products was similar in both composite resins, but differences existed in the amounts of the products released. Both BisHPPP and methacrylic acid were produced in significantly greater amounts as a result of biodegradation of Z250 composite resin compared with TPH. However, more E-BPA was produced as a consequence of TPH degradation, although less than BisHPPP and methacrylic acid by-products. These differences can be explained by the variability of resin composite composition with respect to monomer types, monomer ratios, filler content and filler-to-resin ratios.<sup>11,12,40</sup>

Surface morphology of the cured composite was also found to contribute to the observed differences, as some configurations offer easier access to human salivary esterases and, therefore, more binding sites.<sup>11,12,40</sup> Another factor may be the fact that urethane-modified BisGMA resin associated with TPH is more stable and resistant to hydrolysis and, thus, results in less release of degradation products.<sup>12</sup> Furthermore, dental resin composites with higher silanated filler content have been shown to be less susceptible to the effects of degradation,<sup>11</sup> unlike non-silanated filler particles, which are more susceptible, because of the lack of chemical bonding between the filler and the matrix.<sup>40</sup> Therefore, the degradation product profile varies in terms of identity and amount depending on the composition of the resin composite or adhesive under investigation.

## Biodegradation of the Resin–Dentin Interface

The resin–dentin interface is also susceptible to biodegradation.<sup>25,26,41-44</sup> In a test of the effect of human saliva on totaletch adhesive, Shokati et al.<sup>26</sup> incubated adhesive resin (Scotchbond Multi Purpose, 3M ESPE. St. Paul, MN, USA), resin composite (Z250, St. Paul, MN, USA) and resin composite– dentin mini short-rod specimens with human-saliva-derived esterases (HSDE) for up to 180 days (pH 7.0 and 37°C). The amount of a BisGMA-derived product, BisHPPP (**Fig. 2**), released into the incubation solutions was used as a marker of degradation of resin matrix and polymerized adhesive resin. Because BisGMA is a high molecular weight molecule with rigid phenyl rings and hydrogen bonding capacity, its diffusion through the resin matrix and out to the surface to interact with the enzymes is limited. Therefore, BisHPPP is a good indicator of resin matrix degradation.<sup>10-12,17,32</sup>

This work revealed that HSDE degraded both adhesive resin and resin composite to produce BisHPPP in comparable amounts. The strength of the resin composite–dentin bond was measured as the interfacial fracture toughness of mini short-rod specimens. Specimens incubated for 180 days in ESSENTIAL DENTAL KNOWLEDGE

HSDE had the least resistance to fracture, while non-incubated specimens had the highest resistance. These results indicate that human salivary esterases compromise interfacial integrity by reducing the strength of the bond.<sup>23</sup>

In a more recent investigation, Serkies et al.,<sup>25</sup> found similar effects on total-etch interfaces; however, self-etch specimens showed no reduction in fracture resistance even after 180-day incubation with simulated salivary esterase activities. The authors concluded that material composition and mode of application could affect the biochemical stability of the resin-dentin interface.

In another study, Kermanshahi et al.<sup>23</sup> found an increase in the amount of BisHPPP and interfacial cariogenic bacterial microleakage following incubation of resin composite bonded to dentin using a total-etch system after incubation with salivary esterase-like activity for up to 90 days.

Using transmission electron microscopy, Jung et al.<sup>45</sup> found increased nanoleakage in the resin-dentin interface after storage in an esterase solution compared with a phosphate buffer solution after 4 weeks. Using collagenase and acetylcholinesterase to simulate oral salivary enzymes, Chiaraputt et al.<sup>41</sup> found that the microtensile bond strength of resin composite (Z350, 3M) bonded to human dentin using 4 different total-etch or self-etch adhesives was significantly lower when incubated in enzymes compared with incubation in water or not incubated. They also found that the self-etch adhesives exhibited "water-tree" patterns within the adhesive layer, probably because of the hydrophilicity of the adhesive,<sup>25,26</sup> and that the total-etch adhesive exhibited nanoleakage within the hybrid layer and the adhesive, probably because of its incomplete penetration into the demineralized collagen network.<sup>25,26</sup> Both phenomena compromised the interface, potentially affecting the restoration's longevity.

These findings provide further evidence of biodegradation of the resin-dentin interface caused by salivary enzymes and show that the effect is materials dependent. These studies emphasize that alongside surface resin composite degradation, the resin-dentin interface is also degraded by human or simulated human salivary esterases.<sup>25,26,34,41-44</sup>

#### Biodegradation of Resin Composites and Adhesives by Bacteria and the Reciprocal Effect on the Bacteria

Although studies have investigated the impact of oral saliva on dental resin composites and adhesives, few have looked at the effect of bacteria on these substances. Such research has shown bacteria's ability to penetrate microgaps caused by esterases and form colonies.

Kermanshahi et al.<sup>23</sup> showed that exposing dentincomposite restorations bonded with total-etch adhesive to salivary esterase-like activity resulted in the formation of micro-gaps that were infiltrated and colonized by biofilms of the cariogenic bacteria *Streptococcus mutans*. Dental caries are believed to be the result of acid released by bacterial activity that leads to the demineralization of tooth structures. *Streptococcus mutans* has been found in dental plaque at the margins of composite fillings and is regarded as a major etiological agent responsible for dental caries.

Furthermore, *Streptococcus mutans* has been shown to exhibit esterase activity at levels capable of hydrolytic-mediated degradation of cured dental resin composites and adhesives.<sup>13</sup> Bourbia et al.<sup>13</sup> incubated a standard photopolymerized resin composite (Z250, 3M ESPE, St. Paul, MN, USA) and 2 adhesive resins — a total-etch (Scotchbond, 3M ESPE, St. Paul, MN, USA) and a self-etch (Easybond, 3M ESPE, St. Paul, MN, USA) resin — with the bacteria for up to 30 days. Following incubation, the medium contained the biodegradation by-product BisHPPP and the surface of the resins appeared rougher. The latter finding was in agreement with other investigations that showed an increase in surface degradation of dental resin composites incubated with bacteria (*S. mutans*, *S. gordonii* and *S. sanguis*).<sup>46-48</sup>

Overall, these results point toward the degradative effects of bacteria on dental resin composites and adhesives and highlight the potential for secondary caries and changes in esthetic properties seen clinically with the use of resin materials in dental restorations.

Bacteria-catalyzed biodegradation is further exacerbated by the effect of the by-products from dental resin composites and adhesives on the bacteria. Studies have revealed that by-products, such as TEG, increase bacterial growth and genetic expression of virulence.<sup>7,18</sup> At levels found *in vivo*, TEG modulates the level of expression of glucosyltransferase B (gtfB), which is involved in biofilm formation, and *yfiV*, a putative transcription regulator.<sup>21</sup>

More recent studies have demonstrated a broader effect of TEG and BisHPPP on gene expression, as well as associated proteins related to the virulence of the cariogenic bacteria, *Streptococcus mutans*.<sup>19,20</sup> These findings directly link biodegradation to bacterial proliferation in the oral cavity. This implies that resin composites, in their current form, are not only structurally vulnerable, but can also contribute to the progression of oral disease by increasing growth and virulence of the bacteria that grow over them and within the restoration-tooth margins.

On the other hand, other studies have found that BisGMA degradation products (BisHPPP and methacrylic acid) slightly inhibit growth of *Streptococcus mutans*,<sup>7,21</sup> suggesting that different degradation products have different effects on oral bacteria and that the effect is strain-dependent. Therefore, to reach a convincing conclusion on the complete effect of degradation products on bacterial activity in the oral cavity, research must be conducted on the effect of cumulative degradation products on bacterial growth and virulence, also in multi-species cultures relevant to dental plaque.



# Conclusion

Although dental resin composite and adhesive restoration materials have advantages, such as esthetic appeal, and no perceived danger of mercury poisoning, they seem to be at a disadvantage in terms of long-term stability, as they are affected by physical and biochemical challenges. Biochemical interactions may favour the proliferation of cariogenic bacteria and alter their virulence, thus potentially contributing to an increase in secondary caries and resin composite failure. Because the oral cavity is complex and various influences are acting in concurrence, it is evident that more research must be done to better understand the problems surrounding resin composites and adhesives to improve not only their physical properties, but also their biostability and interactions with oral biofilms.

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# References

- Opdam NJ, Bronkhorst EM, Loomans BA, Huysmans MC. 12-year survival of composite vs. amalgam restorations. J Dent Res. 2010;89(10):1063-7.
- Opdam NJ, van de Sande FH, Bronkhorst E, Cenci MS, Bottenberg P, Pallesen U, et al. Longevity of posterior composite restorations: a systematic review and meta-analysis. <u>J Dent Res. 2014;93(10):943-9</u>.
- Soncini JA, Maserejian NN, Trachtenberg F, Tavares M, Hayes C. The longevity of amalgam versus compomer/ composite restorations in posterior primary and permanent teeth: findings from the New England Children's Amalgam Trial. <u>J Am Dent Assoc. 2007;138(6):763-72</u>.
- Burke FJ, McHugh S, Hall AC, Randall RC, Widstrom E, Forss H. Amalgam and composite use in UK general dental practice in 2001. <u>Br Dent J. 2003;194(11):613-8</u>; discussion 609.

- 5. Burke FJ, Wilson NH, Cheung SW, Mjör IA. Influence of patient factors on age of restorations at failure and reasons for their placement and replacement. <u>J Dent.</u> 2001;29(5):317-24.
- Demarco FF, Corrêa MB, Cenci MS, Moraes RR, Opdam NJ. Longevity of posterior composite restorations: not only a matter of materials. <u>Dent Mater. 2012;28(1):87-101</u>.
- Khalichi P, Cvitkovitch DG, Santerre JP. Effect of composite resin biodegradation products on oral streptococcal growth. <u>Biomaterials. 2004;25(24):5467-72</u>.
- Santerre JP, Shajii L, Tsang H. Biodegradation of commercial dental composites by cholesterol esterase. <u>J Dent Res.</u> <u>1999;78(8):1459-68</u>.
- Santerre JP, Shajii L, Leung BW. Relation of dental composite formulations to their degradation and the release of hydrolyzed polymeric-resin-derived products. <u>Crit</u> <u>Rev Oral Biol Med. 2001;12(2):136-51</u>.
- Finer Y, Santerre JP. Salivary esterase activity and its association with the biodegradation of dental composites. J Dent Res. 2004;83(1):22-6.
- Finer Y, Santerre JP. Influence of silanated filler content on the biodegradation of bisGMA/TEGDMA dental composite resins. <u>J Biomed Mater Res A</u>. 2007;81(1):75-84.
- Finer Y, Santerre JP. The influence of resin chemistry on a dental composite's biodegradation. <u>J Biomed Mater Res A.</u> <u>2004;69(2):233-46</u>.
- Bourbia M, Ma D, Cvitkovitch DG, Santerre JP, Finer Y. Cariogenic bacteria degrade dental resin composites and adhesives. <u>J Dent Res.</u> 2013;92(11):989-94.
- Cai K, Delaviz Y, Banh M, Guo Y, Santerre JP. Biodegradation of composite resin with ester linkages: identifying human salivary enzyme activity with a potential role in the esterolytic process. <u>Dent Mater. 2014;30(8):848-60</u>.
- Delaviz Y, Finer Y, Santerre JP. Biodegradation of resin composites and adhesives by oral bacteria and saliva: a rationale for new material designs that consider the clinical environment and treatment challenges. <u>Dent Mater.</u> <u>2014;30(1):16-32</u>.
- Lin BA, Jaffer F, Duff MD, Tang YW, Santerre JP. Identifying enzyme activities within human saliva which are relevant to dental resin composite biodegradation. <u>Biomaterials.</u> 2005;26(20):4259-64.
- Jaffer F, Finer Y, Santerre JP. Interactions between resin monomers and commercial composite resins with human saliva derived esterases. <u>Biomaterials</u>. 2002;23(7):1707-19.
- Khalichi P, Singh J, Cvitkovitch DG, Santerre JP. The influence of triethylene glycol derived from dental composite resins on the regulation of Streptococcus mutans gene expression. <u>Biomaterials. 2009;30(4):452-9</u>.
- Sadeghinejad L, Cvitkovitch DG, Siqueira WL, Merritt J, Santerre JP, Finer Y. Mechanistic, genomic and proteomic study on the effects of BisGMA-derived biodegradation product on cariogenic bacteria. <u>Dent Mater.</u> 2017;33(2):175-90.
- Sadeghinejad L, Cvitkovitch DG, Siqueira WL, Santerre JP, Finer Y. Triethylene glycol up-regulates virulence-associated genes and proteins in Streptococcus mutans. <u>PloS One.</u> <u>2016;11(11):e0165760</u>.
- Singh J, Khalichi P, Cvitkovitch DG, Santerre JP. Composite resin degradation products from BisGMA monomer modulate the expression of genes associated with biofilm formation and other virulence factors in Streptococcus mutans. <u>J Biomed Mater Res A. 2009;88(2):551-60</u>.

ESSENTIAL DENTAL KNOWLEDGE

- 22. Brännström M. Communication between the oral cavity and the dental pulp associated with restorative treatment. Oper Dent. 1984;9(2):57-68.
- Kermanshahi S, Santerre JP, Cvitkovitch DG, Finer Y. Biodegradation of resin-dentin interfaces increases bacterial microleakage. <u>J Dent Res. 2010;89(9):996-1001</u>.
- Murray PE, Windsor LJ, Smyth TW, Hafez AA, Cox CF. Analysis of pulpal reactions to restorative procedures, materials, pulp capping, and future therapies. <u>Crit Rev Oral Biol Med.</u> 2002;13(6):509-20.
- Serkies KB, Garcha R, Tam LE, De Souza GM, Finer Y. Matrix metalloproteinase inhibitor modulates esterase-catalyzed degradation of resin-dentin interfaces. <u>Dent Mater.</u> <u>2016;32(12):1513-23</u>.
- Shokati B, Tam LE, Santerre JP, Finer Y. Effect of salivary esterase on the integrity and fracture toughness of the dentin-resin interface. <u>J Biomed Mater Res B Appl Biomater</u>. 2010;94(1):230-7.
- Van Meerbeek B, Van Landuyt K, De Munck J, Hashimoto M, Peumans M, Lambrechts P, et al. Techniquesensitivity of contemporary adhesives. <u>Dent Mater J.</u> <u>2005;24(1):1-13</u>.
- 28. Ferracane JL. Resin composite state of the art. <u>Dent.</u> <u>Mater. 2011;27(1):29-38</u>.
- Klapdohr S, Moszner N. New inorganic components for dental filling composites. <u>Monatshefte für Chemie.</u> 2005;136(1):21-45.
- Obici AC, Sinhoreti MAC, Frollini E, Correr Sobrinho L, Consani S. Degree of conversion of Z250 composite determined by fourier transform infrared spectroscopy: comparison of techniques, storage periods and photoactivation methods. <u>Mater Res. 2004;7(4):605-10.</u>
- Finer Y, Jaffer F, Santerre JP. Mutual influence of cholesterol esterase and pseudocholinesterase on the biodegradation of dental composites. <u>Biomaterials. 2004;25(10):1787-93</u>.
- Finer Y, Santerre JP. Biodegradation of a dental composite by esterases: dependence on enzyme concentration and specificity. <u>J Biomater Sci Polym Ed. 2003;14(8):837-49</u>.
- Hagio M, Kawaguchi M, Motokawa W, Miyazaki K. Degradation of methacrylate monomers in human saliva. <u>Dent Mater J. 2006;25(2):241-6</u>.
- Hsu WY, Wang VS, Lai CC, Tsai FJ. Simultaneous determination of components released from dental composite resins in human saliva by liquid chromatography/multiple-stage ion trap mass spectrometry. <u>Electrophoresis</u>. 2012;33(4):719-25.
- Van Landuyt KL, Snauwaert J, De Munck J, Peumans M, Yoshida Y, Poitevin A, et al. Systematic review of the chemical composition of contemporary dental adhesives. <u>Biomaterials. 2007;28(26):3757-85</u>.
- Osorio E, Toledano M, Yamauti M, Osorio R. Differential nanofiller cluster formations in dental adhesive systems. <u>Microsc Res Tech. 2012;75(6):749-57</u>.
- Peumans M, Kanumilli P, De Munck J, Van Landuyt K, Lambrechts P, Van Meerbeek B. Clinical effectiveness of contemporary adhesives: a systematic review of current clinical trials. <u>Dent Mater. 2005;21(9):864-81</u>.
- Moszner N, Salz U, Zimmermann J. Chemical aspects of self-etching enamel-dentin adhesives: a systematic review. Dent Mater. 2005;21(10):895-910.
- Mirmohammadi H, Kleverlaan CJ, Aboushelib MN, Feilzer AJ. Influence of salivary enzymes and alkaline pH environment on fatigue behavior of resin composites. <u>Am J</u> <u>Dent. 2011;24(1);31-6</u>.

- Shajii L, Santerre JP. Effect of filler content on the profile of released biodegradation products in micro-filled bis-GMA/TEGDMA dental composite resins. <u>Biomaterials.</u> <u>1999;20(20):1897-908</u>.
- Chiaraputt S, Roongrujimek P, Sattabanasuk V, Panich N, Harnirattisai C, Senawongse P. Biodegradation of all-in-one self-etch adhesive systems at the resin-dentin interface. <u>Dent Mater J. 2011;30(6):814-26</u>.
- 42. Park JG, Ye Q, Topp EM, Spencer P. Enzyme-catalyzed hydrolysis of dentin adhesives containing a new urethanebased trimethacrylate monomer. <u>J Biomed Mater Res B</u> <u>Appl Biomater. 2009;91(2):562-71</u>.
- Skovron L, Kogeo D, Gordillo LA, Meier MM, Gomes OM, Reis A, et al. Effects of immersion time and frequency of water exchange on durability of etch-and-rinse adhesive. <u>J Biomed Mater Res B Appl Biomater. 2010;95(2):339-46</u>.
- Zou Y, Jessop JL, Armstrong SR. In vitro enzymatic biodegradation of adhesive resin in the hybrid layer. J Biomed Mater Res A. 2010;94(1):187-92.
- Jung YJ, Hyun HK, Kim YJ, Jang KT. Effect of collagenase and esterase on resin-dentin interface: a comparative study between a total-etch adhesive and a self-etch adhesive. <u>Am J Dent. 2009;22(5):295-8</u>.
- Fúcio SB, Carvalho FG, Sobrinho LC, Sinhoreti MA, Puppin-Rontani RM. The influence of 30-day-old Streptococcus mutans biofilm on the surface of esthetic restorative materials — an in vitro study. <u>J Dent. 2008;36(10):833-9</u>.
- Gregson KS, Shih H, Gregory RL. The impact of three strains of oral bacteria on the surface and mechanical properties of a dental resin material. <u>*Clin Oral Investig.* 2012;16(4):1095-103</u>.
- Beyth N, Bahir R, Matalon S, Domb AJ, Weiss El. Streptococcus mutans biofilm changes surface-topography of resin composites. <u>Dent Mater. 2008;24(6):732-6</u>.